

# **9-[(2'S,3'S)-3'-FORMYL-2',3'-DIHYDROXYPROPYL]ADENINE: A FACILE AFFINITY-LABELING PROBE OF HUMAN S-ADENOSYL-L-HOMOCYSTEINE HYDROLASE**

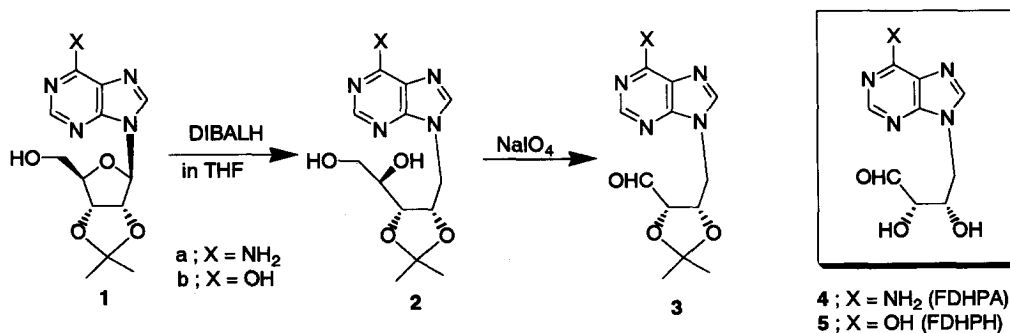
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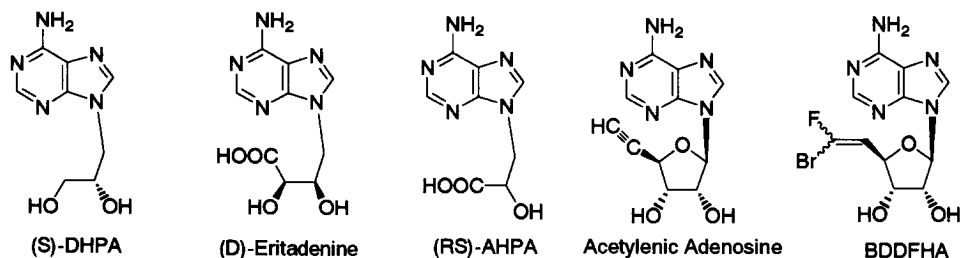
**Abstract:** Treatment of human recombinant *S*-adenosyl-L-homocysteine (SAH) hydrolase with 9-[(2'S,3'S)-3'-formyl-2',3'-dihydroxypropyl]adenine (FDHPA) caused irreversible inactivation in a time- and concentration-dependent manner ( $K_i = 8.8 \mu\text{M}$ ,  $k_{\text{inact}} = 0.09 \text{ min}^{-1}$ ). FDHPA behaved as a facile affinity-labeling probe of SAH hydrolase. © 1999 Elsevier Science Ltd. All rights reserved.

The cellular enzyme *S*-adenosyl-L-homocysteine (SAH) hydrolase (EC 3.3.1.1) has emerged as a target enzyme for the molecular design of anti-viral agents.<sup>1</sup> Inhibition of SAH hydrolase results in cellular accumulation of SAH, which is a potent product inhibitor of *S*-adenosyl-L-methionine-dependent biological methylation. We have recently found a convenient method for the direct preparation of 9-ribitylpurines (**2**) by the reductive cleavage of purine nucleosides (**1**) with diisobutylaluminum hydride (DIBALH).<sup>2</sup> Various acyclic nucleosides such as (*S*)-9-(2,3-dihydroxypropyl)adenine [(*S*)-DHPA],<sup>4</sup> (D)-critadenine,<sup>5</sup> and (*RS*)-3-adenin-9-yl-2-hydroxypropanoic acid [(*RS*)-AHPA]<sup>6</sup> have been found to exhibit a broad spectrum of antiviral



Scheme 1

properties due to the inhibition of SAH hydrolase. Affinity-labeling probes such as acetylenic adenosine<sup>3a</sup> and BDDFHA<sup>3b,c</sup> were prepared for the elucidation of the molecular mechanism of SAH hydrolase (Scheme 2).



**Scheme 2**

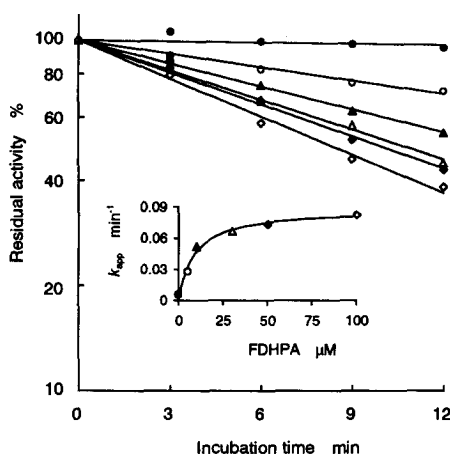
During the course of studies on the preparation of biologically important nucleosides<sup>7,8</sup> using the reductive cleavage of purine nucleosides,<sup>2</sup> a DHPA analogue possessing a formyl group at the 3'-position has been designed as a possible affinity-labeling probe for the elucidation of the catalytic site of SAH hydrolase. In this paper, we describe a method for the preparation of a facile affinity-labeling probe and its biological properties against human recombinant SAH hydrolase.

Thus, the possible affinity-labeling probe, 9-[(2'S,3'S)-3'-formyl-2',3'-dihydroxypropyl]adenine (**4**; FDHPA), was prepared from naturally-occurring nucleosides (**1**) *via* reductive cleavage with DIBALH (Scheme 1). Subsequent oxidation of **2** with NaIO<sub>4</sub> gave the corresponding formyl derivatives (**3**) quantitatively. Careful deprotection of **3** with 2% trifluoroacetic acid gave FDHPA (**4**) in good yield.<sup>9</sup>

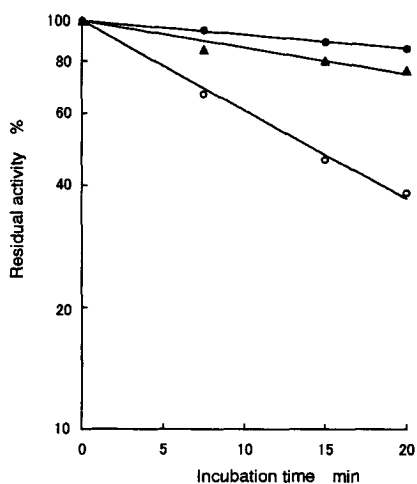
The cDNA<sup>10</sup> for human SAH hydrolase has been cloned from human hepatoma cell HepG2 by RT-PCR and the obtained human recombinant SAH hydrolase was purified according to the literature.<sup>11</sup> Incubation with FDHPA (**4**) caused inactivation of the human SAH hydrolase and the activity was not recovered by dialysis. The inactivation was followed by the pseudo-first-order reaction (Figure 1). However,  $k_{app}$  increased in a dose-dependent manner and reached maximum at 100  $\mu$ M FDHPA (**4**) (Figure 1, inset). The values of  $K_i$  and  $k_{inact}$ , which are useful values to evaluate the affinity and reactivity of an affinity-labeling reagent,<sup>3c,12</sup> were 8.8  $\mu$ M and 0.09 min<sup>-1</sup>, respectively.

To elucidate the FDHPA acts as an affinity-labeling probe, a hypoxanthine derivative of FDHPA (**4**), 9-[(2'S,3'S)-3'-formyl-2',3'-dihydroxypropyl]hypoxanthine (**5**; FDHPH), was also synthesized and compared. Despite its structural similarity with FDHPA (**4**), FDHPH (**5**) showed only minimal inactivation effect (Figure 2). Furthermore, SAH hydrolase activity was completely protected from the inactivation by FDHPA (**4**) with

co-existence of 1 mM adenosine (data not shown).



**Figure 1.** Concentration-dependent inactivation of SAH hydrolase by FDHPA (4) SAH hydrolase was incubated with or without each concentration of FDHPA (4) at 30°C (0 μM (●), 5 μM (○), 10 μM (△), 30 μM (▲), 50 μM (◆), 100 μM (◇)). Aliquots of the reaction mixture were removed for evaluating the enzymatic activity at indicated intervals. Inset indicates the secondary plot of FDHPA (4) concentration versus apparent pseudo-first-order rate of inactivation.



**Figure 2.** Inhibitory effect of FDHPA (4) on the SAH hydrolase activity SAH hydrolase was incubated under the absence (●) or presence of 0.1 mM FDHPA (4) (▲) and 0.1 mM FDHPH (5) (△) at 30°C.

Taking the above results into consideration, it is deduced that FDHPA (4) functions as a useful affinity-

labeling probe of SAH hydrolase and provides a clue to elucidation of the molecular mechanism of SAH hydrolase aiming at the molecular design of anti-viral drugs.

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9. The detailed synthetic method for the preparation of **4** and **5** will be reported separately.
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11. a) Gomi, T.; Date, T.; Ogawa, H.; Fujioka, M.; Aksamit, R.R.; Backlund, P.S., Jr.; Cantoni, G.L. *J. Biol. Chem.* **1995**, *270*, 16140. b) The  $V_{max}$  and  $K_m$  values of the human recombinant SAH hydrolase to adenosine were 3.4  $\mu\text{mol}/\text{min}/\text{mg}$  and 1.6  $\mu\text{M}$ , respectively.
12. The data were analyzed by Kitz and Wilson plot, fitting to the following equation:  $k_{app} = k_{inact} [I] / (K_i + [I])$  in this equation  $k_{app}$ ,  $k_{inact}$ ,  $K_i$  and  $I$  mean pseudo-first-order rate of inactivation, maximum rate of inactivation, inhibition constant and concentration of FDHPA (**4**), respectively. The value of  $k_{app}$  was determined from a plot of the residual activity versus incubation time. The values of  $K_i$  and  $k_{inact}$  were obtained using a curve-fitting program CurveExpert. The proposed equilibrium is as follows:  $\text{I-CHO} + \text{H}_2\text{N-E} \longrightarrow \text{I-CH=N-E}$ . I-CHO and E-NH<sub>2</sub> mean FDHPA (**4**) and SAH hydrolase, respectively.